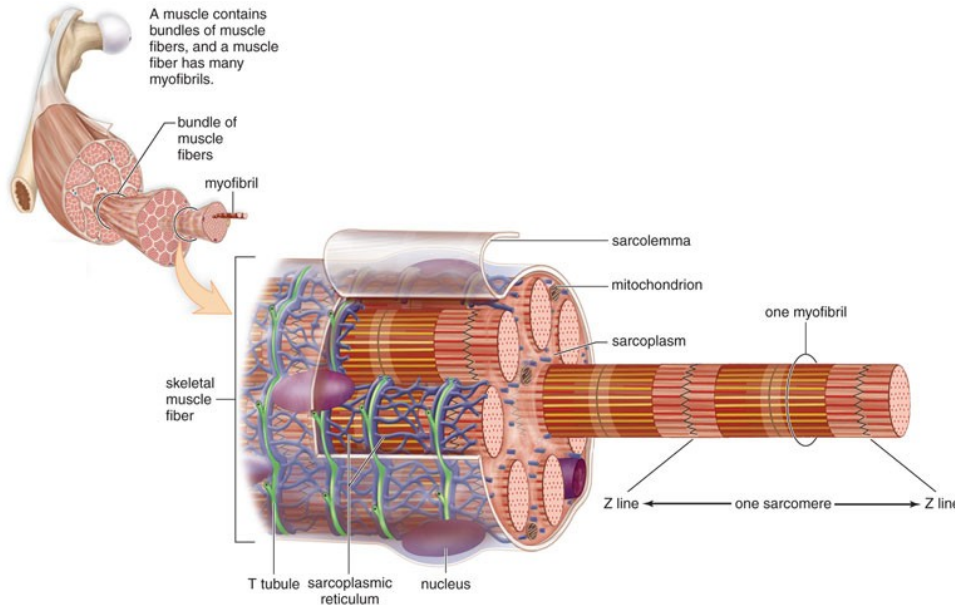


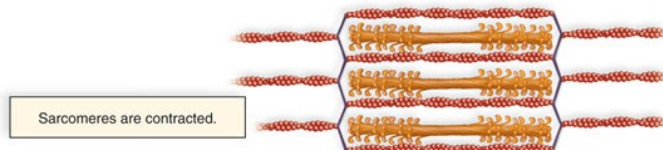
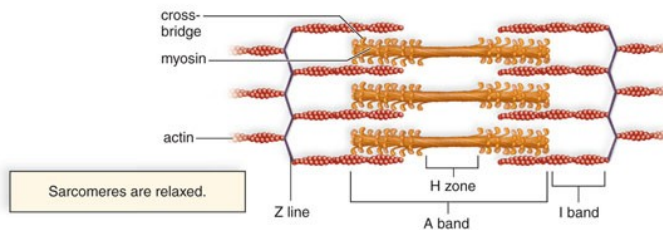
The Equatorial Pattern from Muscle

Myofibrils and Sarcomeres

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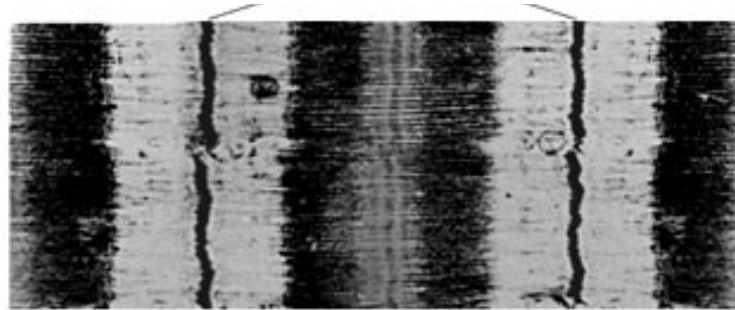
A myofibril has many sarcomeres.



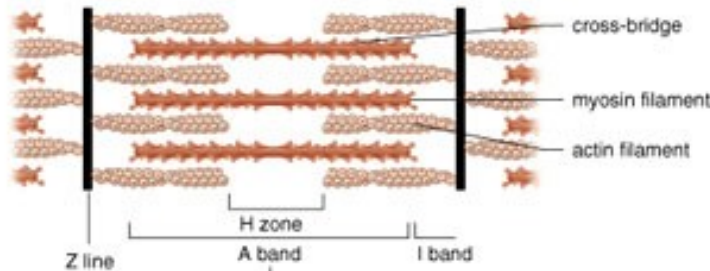
- A myofibril consists of many 2-3 micron long sarcomeres laid end to end
- In the light microscope, sarcomeres show a banding pattern (striations)
- A-band, I band, Z-line, and M-line, H-zone
- Underlying structure can be seen only at electron microscope level

Sarcomeres

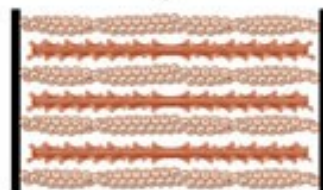
Myofibril has many sarcomeres.



Sarcomere is relaxed.

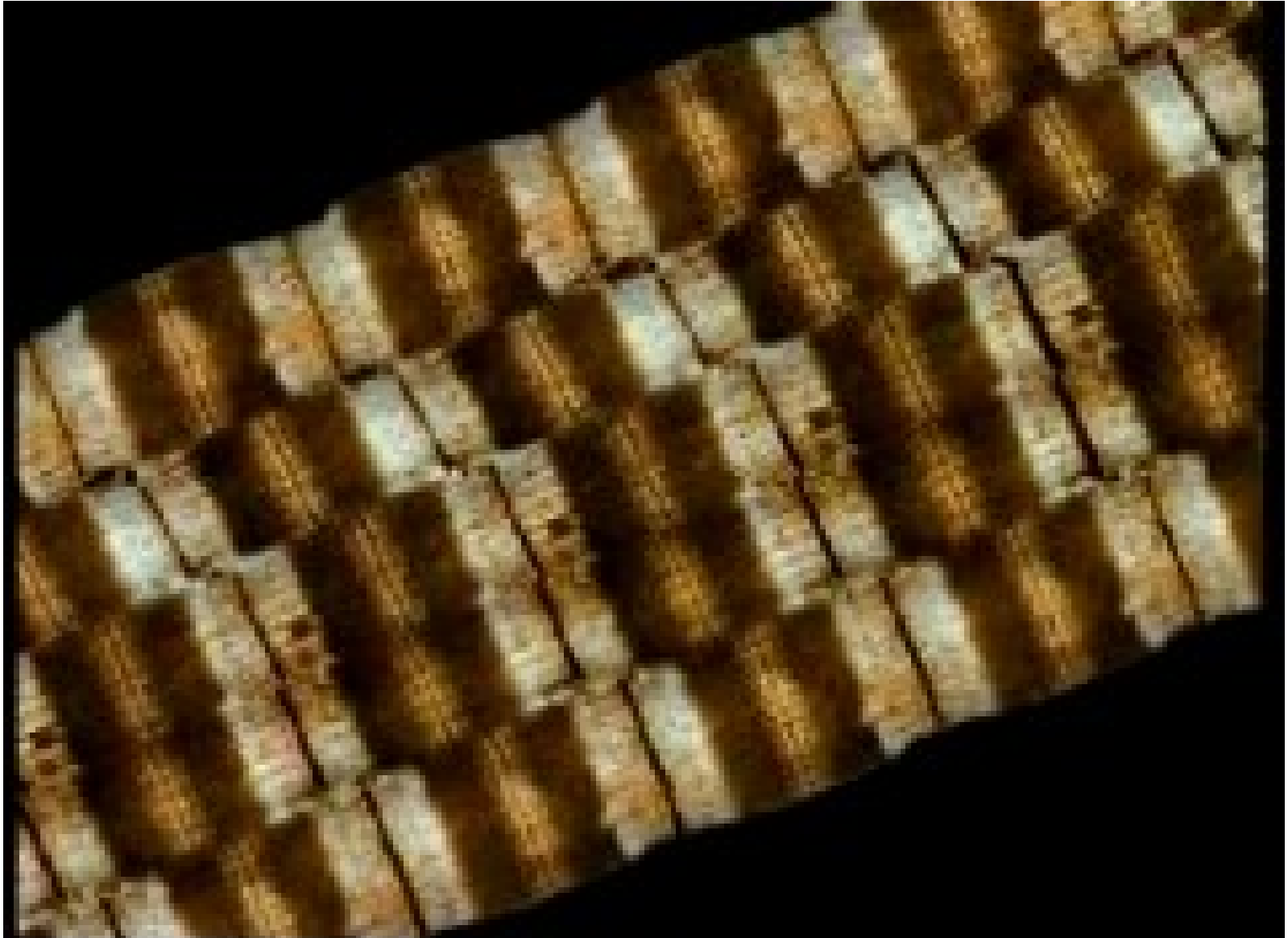


Sarcomere is contracted.

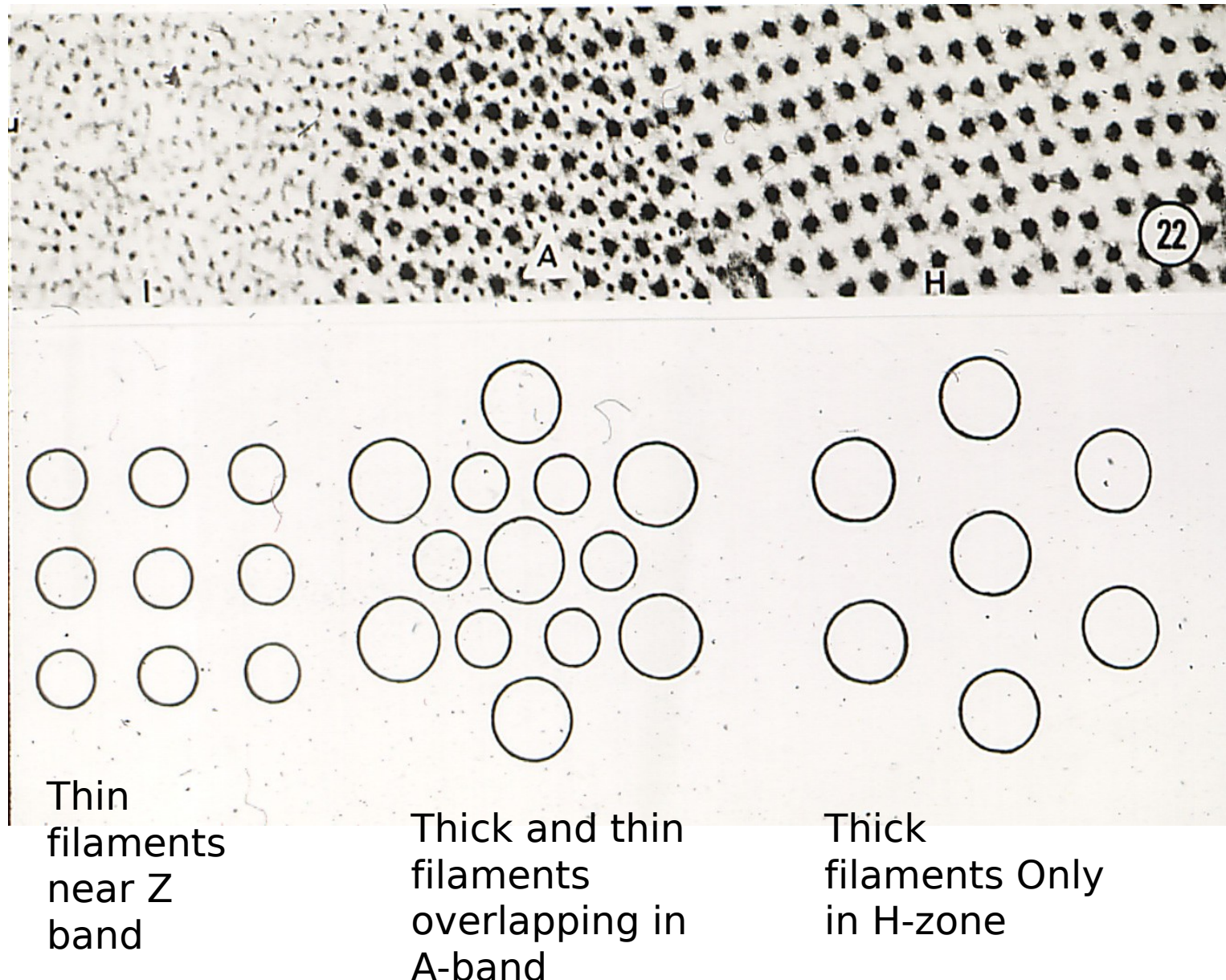


- Sarcomere consists of actin containing thin filaments
- Myosin containing thick filaments
- I band contains only thin filaments
- H-zone only thick
- A-band both thick and thin
- Projections on the thick filament (crossbridges) interact with the thin filament and cause sarcomeres to shorten

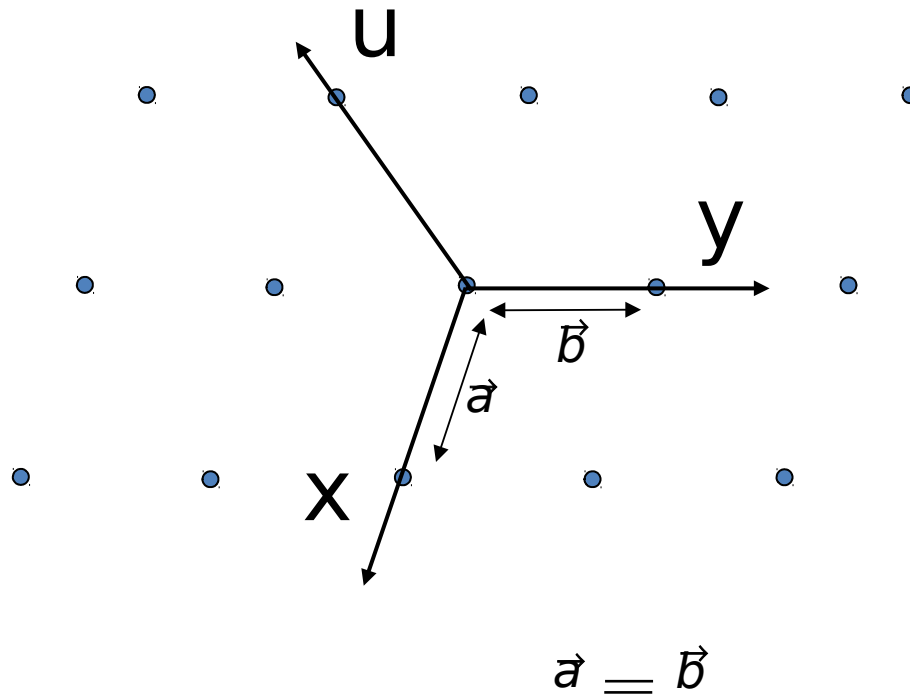
“Sliding Filaments”



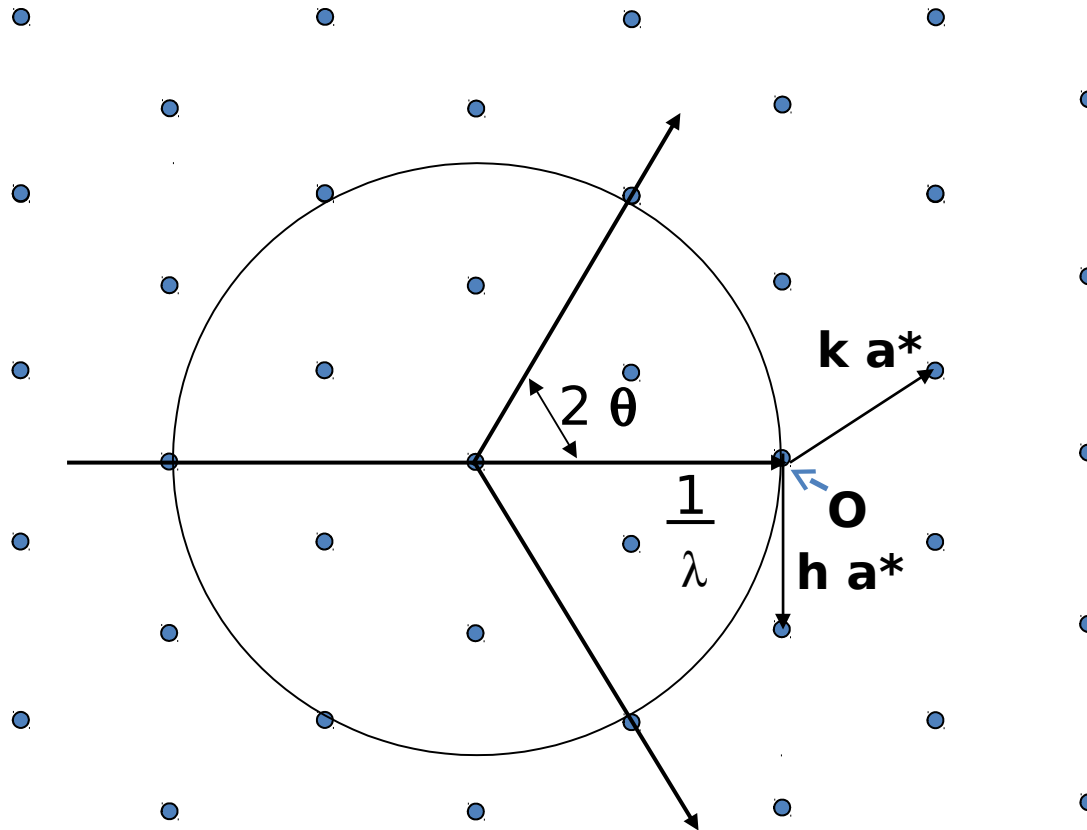
Cross-sections of a sarcomere showing 2-D crystalline structure



Hexagonal Lattice



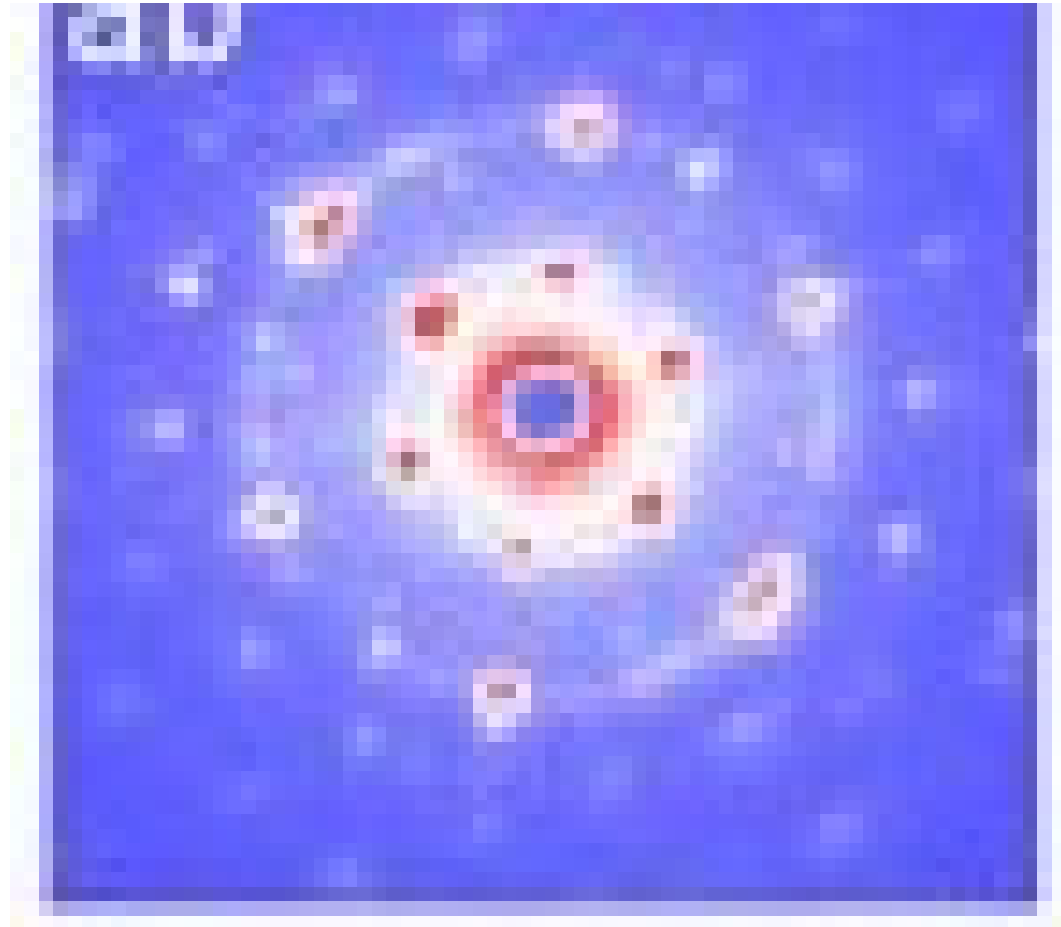
Ewald Sphere



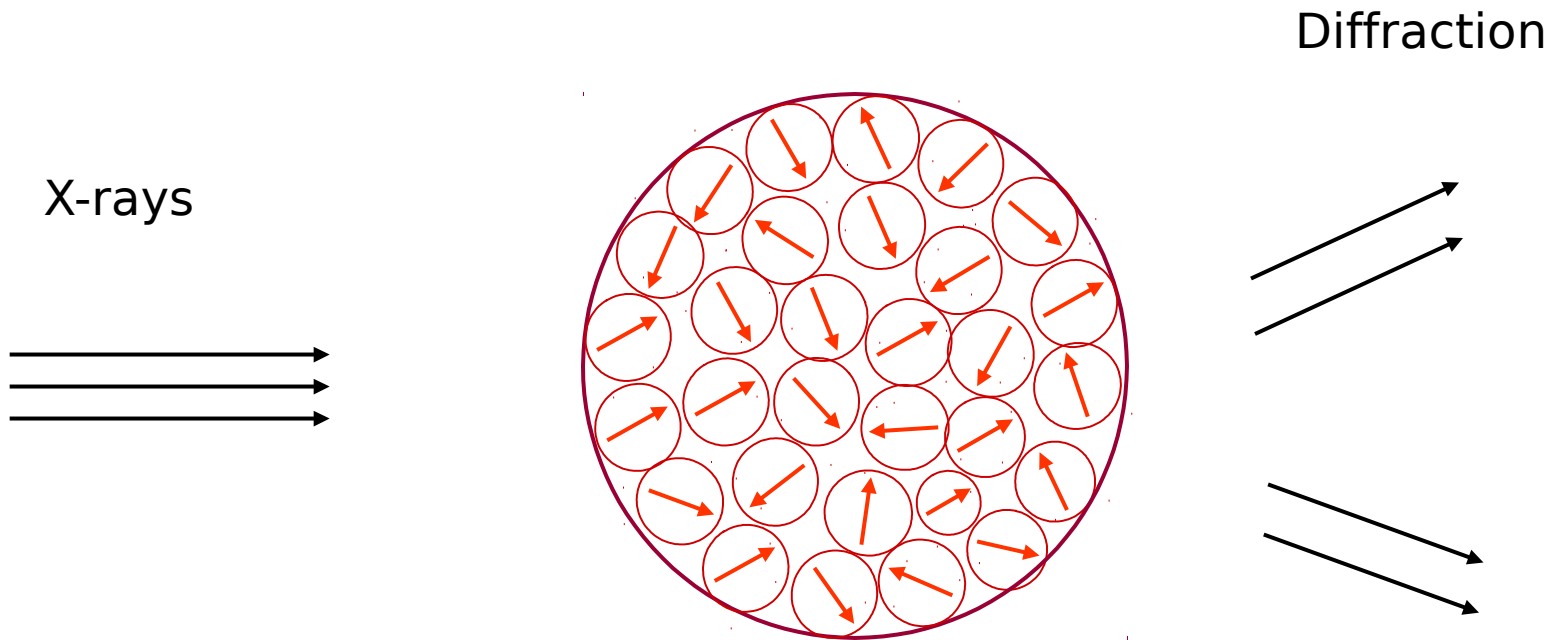
For every lattice in real space there is a corresponding lattice in “reciprocal” i.e. diffraction space. Distances between lattice points are proportional to $1/\text{lattice dimensions in real space}$. The origin of the reciprocal lattice is at O . Lattice points (corresponding to diffraction spots one can observe) are indexed by Miller indices h and k

End on view of hexagonal reciprocal lattice

This was taken by shining the X-ray beam down the axis of a muscle myofibril. (one crystallite)
Not the way we normally do it.



Fiber Cross-section



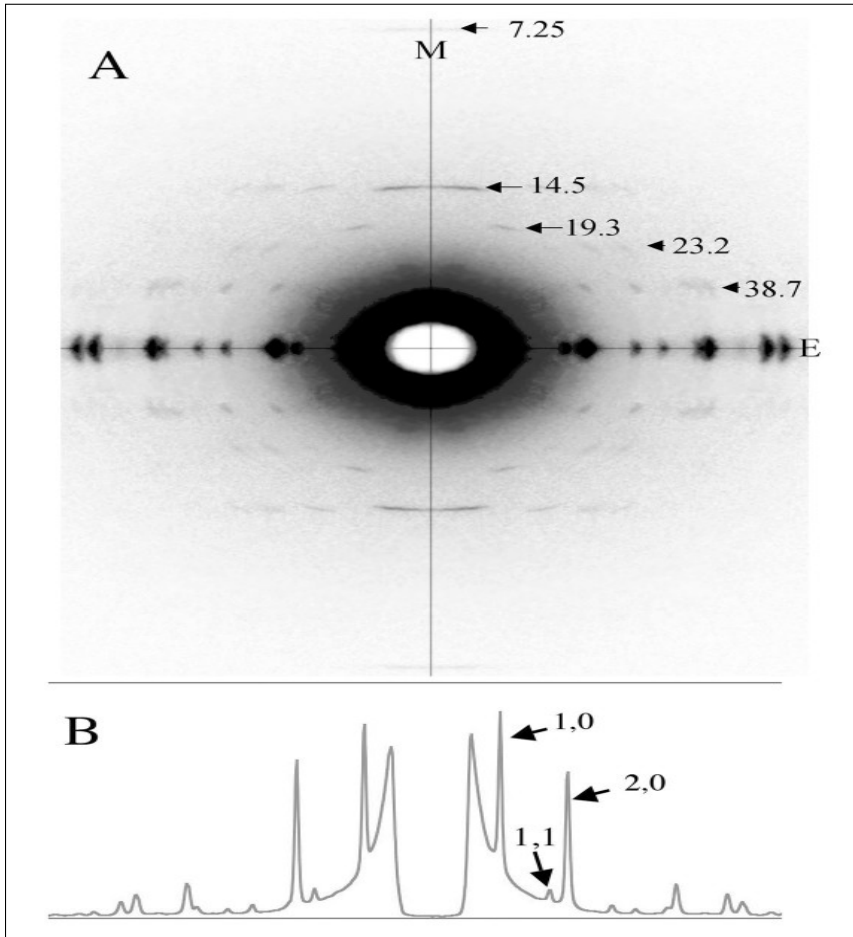
Myofibrils
are at
random
orientatio
ns around
the long
axis of the
muscle
fiber

Complete Statistical rotation:

$$1,0 = 1,0 = 0,1 = 0,-1$$

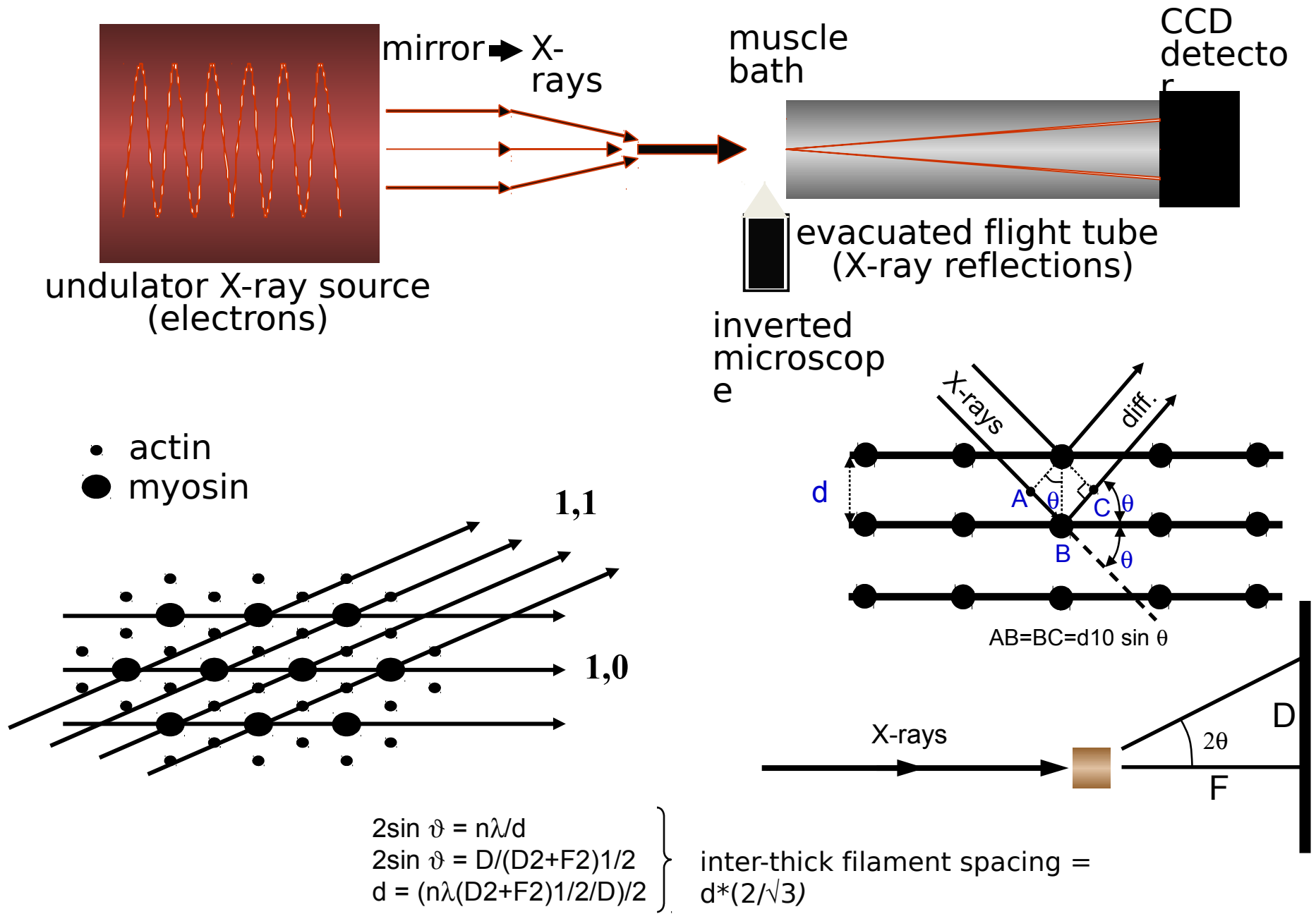
$$1,1 = 1,1 = 1,1 = 1,-1$$

Equatorial pattern from insect muscle



We are looking at projection of hexagonal lattice onto a plane. Insect muscle is highly ordered so you get lots of sharp spots along the equator. Note that they vary in intensity

Experimental arrangement of the BioCAT undulator beam line for X-ray diffraction



Calculating d₁₀

Braggs Law

$$n\lambda = 2d\sin\theta$$

θ is the Bragg angle where 2θ is the angle between the diffracted and incident beam

At small angles

$$\theta = D/2L \text{ so that}$$

$$n\lambda = 2dD/2L$$

Or

$$d = n\lambda L/D$$

So d , the spacing between the diffracting planes is inversely proportional to D , the distance from the origin of the diffraction pattern to a diffraction spot

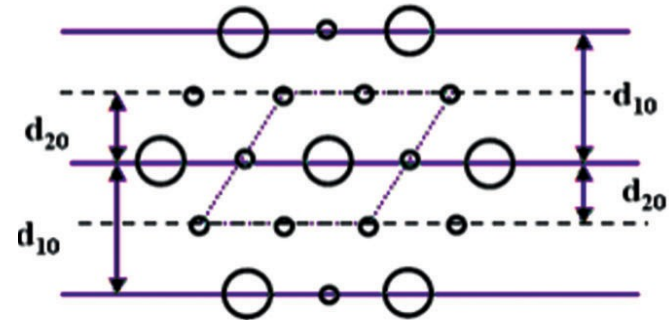
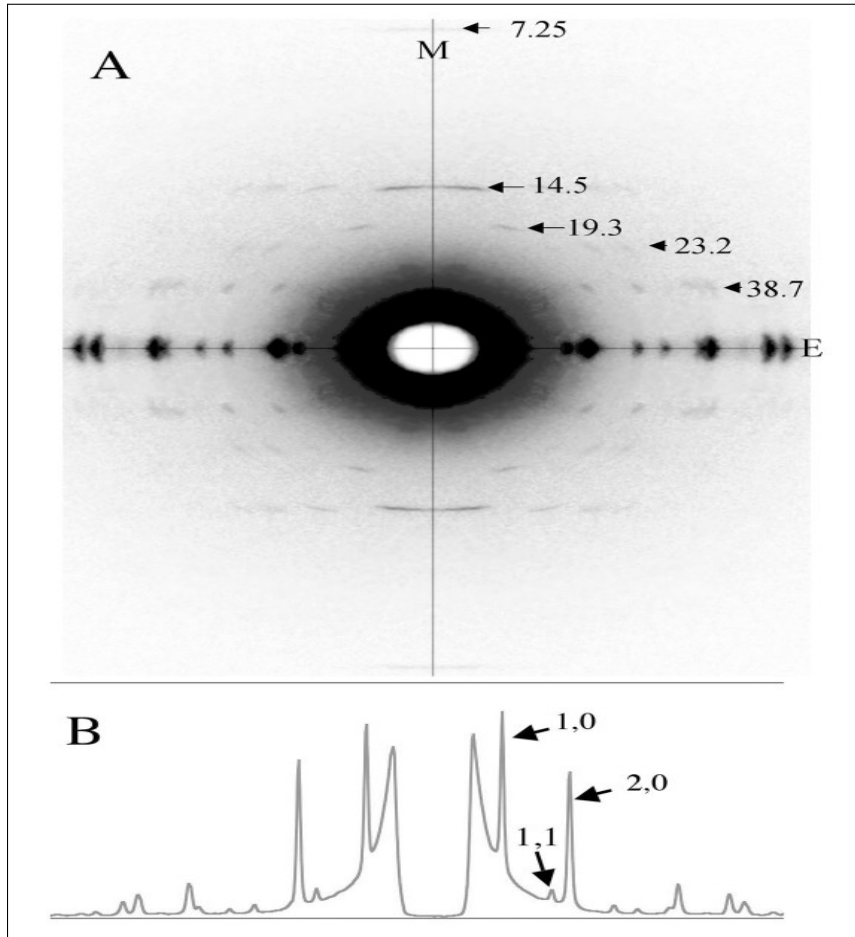
Hexagonal pattern selection rule

- The distances from the center of the pattern to each of the outer reflections ($S_{h,k}$) are related to the distance from the center to the first strong 1,0 reflection, S_{10} , by $S_{h,k} = S_{10}\sqrt{h^2 + k^2 + hk}$ where h and k are the Miller indices of each reflection. Notice that several combinations of h and k values will give rise to the same $S_{h,k}$ meaning that X-ray reflections will superimpose.

Estimating lattice disorder parameters from peak widths

- The width of the Gaussian representing a given diffraction peak $\sigma_{h,k}$ can be expressed as
- $\sigma_{h,k} = \sqrt{(\sigma_c^2 + \sigma_d^2 S_{hk} + \sigma_s^2 S_{hk}^2)}$ where $S_{hk} = \sqrt{h^2 + k^2 + hk}$. σ_c is the known width of the X-ray beam, σ_d is related to the amount of heterogeneity in inter-filament spacing among the myofibrils, and σ_s is related to the amount of paracrystalline (liquid-like) disorder of the myofilaments in the hexagonal lattice.
- These are all interesting physiological parameters

Equatorial Intensities



If crossbridges move away from the thick filament backbone towards the thin filament

Mass leaves the 1,0 plane and joins 2,0

$I_{2,0}/I_{1,0}$ goes up

Use $I_{2,0}/I_{1,0}$ as a measure of degree of association crossbridges with thin filament